In the Specification

Please substitute the following paragraph on page 38, beginning at line 14:

The amount of deprotection of the RNA as assayed using an *in vitro* transcript labelled with 32P and a 5% acrylamide sequencing gel. The rate of migration is proportional to the amount of deprotection and sequencing gels serve as a useful measure of the amount of deprotection that has occured. It was found that with ethanolamine at 55, 45 or 37°C there was significant degradation of the RNA, whilst with ethylenediamine at 55°C there was limited degradation, both deprotection at 45 or 37°C did not lead to detectable RNA degradation. ~~Thgerefore~~ Therefore deprotection with ethanolamine is best achived at 25°C whilst with ethylenediamine, deprotection can be carried out up to 45°C.

Please substitute the following paragraph on page 41, beginning at line 3:

Protected or deprotected RNA may also be separated from hydroxylapatite by inserting 2 wires (anode and cathode) without them touching into a tube containing beads in ~~100□l~~ 100 µl of water or buffer and applying a low voltage such as 5-50V for 5 minutes. The RNA can be recuperated from the liquid phase.

Please substitute the following paragraph on page 41, beginning at line 21 through to page 42, line 5:

Reverse Transcription. 25 ng of the deprotected BMV RNA was added to a 20 µl reaction mixture containing the following final component concentrations: 200 mM Tris-HCl (pH 8.4 at 24°C), 75 mM KCl, 2.5 mM $MgCl_2$, 10 mM DTT, 1 mM dNTP's, 60 ng of oligonucleotide primer BMV R (GAGCCCCAGCGCACTCGGTC) (SEQ ID NO: 1) and MULV RNase $H^-$ (Promega, cat no. M3682, USA). Water was used to bring the final volume to 20 µl. The reaction was allowed to proceed for 20 minutes at 37°C, 20 minutes at 42°C and 20 minutes at 50°C. PCR Amplification. The PCR was carried out in a final volume of 25µl with final concentration of 15mM Tris-HCl pH 8.8, 60mM KCl, 2.5mM $MgCl_2$, 400 µM each dNTP, 10 pmol of each primer BMV F (CTATCACCAAGATGTCTTCG) (SEQ ID NO: 2) and BMV R and 1 unit *Taq* DNA polymerase

(Roche Molecular, France). To the PCR mix was added 2 µl of cDNA generated from the deprotected BMV RNA. Cycle parameters were 94°C x 8 sec, 58°C x 8 sec and 72°C x 15 sec for 30 cycles. The 250bp PCR products were visualised following gel electrophoresis and staining with EtBr.

Please substitute the following paragraph on page 42, beginning at line 24 through to page 43, line 8:

Immobilisation of deprotected RNA. 100, 50 or 25 ng of the deprotected BMV RNA was added to a 20 µl reaction mixture containing the following final component concentrations: 200 mM Tris-HCl (pH 8.4 at 24°C), 75 mM KCl, 2.5 mM $MgCl_2$, 10 mM DTT, 1 mM dNTP's, 60 ng of oligonucleotide primer BMV R (GAGCCCCAGCGCACTCGGTC) (SEQ ID NO: 1) and MULV RNase $H^-$ (Promega, cat no. M3682, USA). Water was used to bring the final volume to 20 µl. The reaction was allowed to proceed for 20 minutes at 37°C, 20 minutes at 42°C and 20 minutes at 50°C. PCR Amplification. The PCR was carried out in a final volume of 25 µl with final concentration of 15mM Tris-HCl pH 8.8, 60mM KCl, 2.5mM $MgCl_2$, ~~400 □M~~ 400mM each dNTP, 10 pmol of each primer BMV F (CTATCACCAAGATGTCTTCG) (SEQ ID NO: 2) and BMV R and 1 unit *Taq* DNA polymerase (Roche Molecular, France). To the PCR mix was added 2 µl of cDNA generated from the deprotected BMV RNA. Cycle parameters were 94°C x 8 sec, 58°C x 8 sec and 72°C x 15 sec for 30 cycles. The 250bp PCR products were visualised following gel electrophoresis and staining with EtBr.

Please insert the Sequence Listing as new page 1 attached hereto following page 60.